

Identifying new susceptibility genes on dopaminergic and serotonergic pathways for the framing effect in decision-making

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Abstract

The framing effect refers the tendency to be risk-averse when options are presented positively but be risk-seeking when the same options are presented negatively during decision-making. This effect has been found to be modulated by the serotonin transporter gene (*SLC6A4*) and the catechol-o-methyltransferase gene (*COMT*) polymorphisms, which are on the dopaminergic and serotonergic pathways and which are associated with affective processing. The current study aimed to identify new genetic variations of genes on dopaminergic and serotonergic pathways that may contribute to individual differences in the susceptibility to framing. Using genome-wide association data and the gene-based principal components regression method, we examined genetic variations of 26 genes on the pathways in 1317 Chinese Han participants. Consistent with previous studies, we found that the genetic variations of the *SLC6A4* gene and the *COMT* gene were associated with the framing effect. More importantly, we demonstrated that the genetic variations of the aromatic-L-amino-acid decarboxylase (*DDC*) gene, which is involved in the synthesis of both dopamine and serotonin, contributed to individual differences in the susceptibility to framing. Our findings shed light on the understanding of the genetic basis of affective decision-making.

Key words: framing effect; decision-making; *DDC*; *COMT*; *SLC6A4*; GWAS

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Introduction

During decision-making, individuals tend to be risk-averse when options are presented in a positive way (i.e. the gain frame) but be risk-seeking when the same options are presented negatively (i.e. the loss frame), a phenomenon known as the 'framing effect' (Tversky and Kahneman, 1981; Kahneman and Tversky, 1984; Kuhberger et al., 1999). This spontaneous bias is observed across different cultures (Kahneman and Tversky, 1979; Sharp and Salter, 1997), and has profound influences on important daily decisions, such as those related to finance, voting, and whether or not to undergo a certain surgery (McNeil et al., 1982; Druckman, 2004).

Previous studies suggested that emotional arousal towards the potential of loss plays an important role in the framing effect. Specifically, psychophysiological evidence demonstrated that choices in the loss frame are associated with more elevated skin conductance responses than choices in the gain frame in normal participants; this effect was absent for autistic participants with emotional impairment (Hill et al., 2004; De Martino et al., 2008). Neuroimaging studies revealed an increased activation of the emotion system (e.g. the amygdala) when participants chose risky options in the loss frame and safe options in the gain frame (De Martino et al., 2006; Roiser et al., 2009; Xu et al., 2013; Gao et al., 2016). Moreover, increased distress results in an increased framing effect (Druckman and McDermott, 2008), while reduced emotional response via cognitive reappraisal decreases individuals' susceptibility to framing (Miu and Crişan, 2011).

The susceptibility to framing in decision-making, which varies substantially across individuals (Kahneman and Tversky, 1979; Sharp and Salter, 1997; De Martino et al., 2006; Roiser et al., 2009; Gao et al., 2016), has moderate heritability (Simonson and Sela, 2011; Cesarini et al., 2012; Cronqvist and Siegel, 2012), suggesting that genetic variations contribute to the individual differences. Although genetic studies on risk-taking have demonstrated the important role of genetic variations on dopaminergic and serotonergic pathways in decision-making under risks (Crişan et al., 2009; Dreber et al., 2009; Kuhnen and Chiao, 2009; He et al., 2010; Frydman et al., 2011; Heitland et al., 2012; Reuter et al., 2013; Set et al., 2014), only a few studies investigated directly the genetic basis of the susceptibility to framing in decision-making. Two studies (Crişan et al., 2009, N = 36; Roiser et al., 2009, N = 30) showed the association between 5-HTTLPR variable number of tandem repeats variation, the genetic variation in the promoter region of the serotonin transporter gene (SLC6A4), and individuals' susceptibility to framing. Individuals who are homozygous for the short (s) allele at the 5-HTTLPR are more susceptible to framing than individuals who are homozygous for the long (l) allele. Our recent work (N = 98) on dopamine degradation gene COMT indicated that COMT Val158Met polymorphism is also associated with the individual differences in susceptibility to framing. Compared with the Val/Val homozygotes, the framing effect is more profound in the Met allele carriers, who have increased prefrontal dopamine concentrations (Gao et al., 2016). However, given that complex molecular networks and cellular pathways contribute to individual differences in complex behaviors (Schadt, 2009; Wang et al., 2010), variations in a few polymorphisms are unlikely to be the whole story behind the genetic basis of susceptibility to framing. Here we sought to further shed light on this genetic basis by investigating variations in a set of genes on the dopaminergic and serotonergic pathways.

Genetic variations related to dopamine metabolism and signaling have been shown to be involved in emotional processing

and dysregulation (Meyer-Lindenberg, 2010; Opmeer et al., 2010; Scharinger et al., 2010; Gadow et al., 2014). For example, the Met allele of the COMT gene is associated with the negative bias in affective processing, including decreased resilience in response to negative mood states and increased anxiety levels and limbic reactivity (e.g. amygdala) in response to unpleasant stimuli (Ohara et al., 1998; Enoch et al., 2003; Schupp et al., 2003; McGrath et al., 2004; Smolka et al., 2005; Drabant et al., 2006; Kia-Keating et al., 2007; Olsson et al., 2007; Montag et al., 2008; Williams et al., 2010; for a review, see Heinz and Smolka, 2006). Similarly, genetic variations of the monoamine oxidase gene (MAOA and MAOB) are key candidates in studies concerning the mechanisms of negative emotionality (Dlugos et al., 2009) and psychiatric disorders [e.g. major depression (Kersting et al., 2007; Roohi et al., 2009)].

Genetic association studies also suggest the contribution of gene variations in the serotonin system to affective processing and psychiatric disorders (Meyer-Lindenberg, 2010; Scharinger et al., 2010; Bevilacqua and Goldman, 2011; Fabbri et al., 2013; Jonassen and Landrø, 2014). Specifically, relative to the homozygous long variation (l/l), the short (s) allele of 5-HTTLPR exhibits increased amygdala reactivity to negative environmental stimuli (Hariri et al., 2002; Canli et al., 2005; Heinz et al., 2005) and to negative self-reflection (Ma et al., 2014); it is thus recognized as a risk allele for affective disorders (Bellivier et al., 1998; Lotrich and Pollock, 2004; Lasky-Su et al., 2005; Uher and McGuffin, 2010), although a meta-analysis suggested that the main effect of 5-HTTLPR genotype and the interaction between 5-HTTLPR and SLE on risk of depression are negligible (Munafò et al., 2009). Moreover, a gene involved in the synthesis of both dopamine and serotonin, the aromatic-L-amino-acid decarboxylase gene (DDC), is associated with affective disorders, including anxiety state (Costas et al., 2010) and bipolar disorder (Børglum et al., 2003).

Considering both the key role of affective processing in the framing effect and the association between emotional processing and genetic variations on dopaminergic and serotonergic pathways, genes on these two pathways are possible contributors to the individual differences in susceptibility to framing. In the current study, by applying the gene-based principal components regression (PCReg) approach (Wang and Abbott, 2008; Hibar et al., 2011a, b) and pathway-based approach (Wang et al., 2007; Wang et al., 2010) to genome-wide association (GWA) data, we sought to find new genes that contribute to the susceptibility to framing in decision-making. To this end, 26 genes were selected from dopaminergic and serotonergic pathways and subjected to analysis. All genes were selected according to dopaminergic pathway and serotonergic pathway maps in KEGG database (<http://www.genome.jp/kegg/>), a manually curated collection of pathway maps widely used in gene-set analysis (e.g. Nemoda et al., 2011; Set et al., 2014; Baou et al., 2016). Genes that are lack of expression in the brain were excluded in the current study.

Given the availability of densely spaced single nucleotide polymorphisms (SNPs) within a gene in GWA data, the traditional SNP-based approach suffers from the collinearity arisen from linkage disequilibrium (LD) among SNPs (Wang and Abbott, 2008) and lacks the power to uncover the relatively small effect sizes conferred by most genetic variations (Wang et al., 2010). To address these issues, the gene-based PCReg method takes into account the common variation within a candidate gene jointly by using a few uncorrelated principal components (PCs) computed from the sample covariance matrix of all SNPs (Neale and Sham, 2004; Wang and Abbott, 2008). This

approach reduces dimensionality of the genetic information and the number of tests, which in turn helps to reduce the problem of chance findings (i.e. false positives) due to multiple testing (Neale and Sham, 2004; Klei et al., 2008; Wang and Abbott, 2008). Compared with the SNP-based approach, the gene-based approach is more efficient when there is weak but coordinated effects arising from multiple SNP markers (Wang et al., 2010; Set et al., 2014) and has been widely used in behavioral genetic and neuroimaging studies (Wang and Abbott, 2008; Hibar et al., 2011a,b).

Materials and methods

Participants

Participants were all incoming freshman (Grade 2013) at Chongqing University of Medical Sciences, China, and were recruited from the freshman seminar as they arrived at university. One thousand five hundred and eighty-two unrelated Chinese Han students (80.1% females, mean age 18.66 ± 0.90 years) were recruited. Participants were divided into 15 cohorts. About 100 participants in the same cohort came to a testing room at the same time, completed the behavioral task on computers and submitted their data to the server. Two hundred and sixty-five of them were excluded from data analysis because of their low accuracy in the catch condition in which they were expected to choose the option with an expected value much higher than the other option, indicating a high probability that they did not actively engage in the task (see the later behavioral test; De Martino et al., 2006; Gao et al., 2016). In all the 1582 participants, 5 participants reported a history of psychiatric, neurological or cognitive disorders in the self-reported questionnaire. These five persons also performed badly in the catch condition and were hence excluded.

A final sample of 1317 participants was included in the following analysis. None of the participants reported any history of psychiatric, neurological or cognitive disorders in the self-reported questionnaire (see Supplementary data for more details about the self-reported questionnaire). All of them were in the normal range of anxiety symptoms (i.e. scores < 50 , mean = 30.58, SD = 5.35) as assessed by the Zung Self-Rating Anxiety Scale (Zung, 1971; Wang et al., 1999) and in the normal range of depressive symptoms (i.e. scores < 50 , mean = 33.3, SD = 6.31) as assessed by the Zung Self-Rating Depression Scale (Zung, 1965; Wang et al., 1999), except for nine participants who had higher scores (51, 51, 53, 53, 53, 54, 54, 55 and 59, respectively) beyond the normal range of depressive symptoms and three participants who had higher scores (51, 51 and 56, respectively) beyond the normal range of anxiety symptoms. Given that excluding these 12 participants did not change the pattern of results, we included them in the following reported data analysis. Written informed consents were obtained from each participant. This study was performed in accordance with the Declaration of Helsinki and was approved by the Ethics Committee of the School of Psychological and Cognitive Sciences, Peking University.

The behavioral test

We used the same behavioral task in Gao et al. (2016), which is developed by De Martino et al (2006) and has been used to assess the framing effect (Roiser et al., 2009; Xu et al., 2013). At the beginning of each trial, participants were endowed with an initial amount of monetary reward. Then they chose between receiving a certain guaranteed amount of money from the initial

amount (i.e. the sure option) and taking a risky option that could enable them, with a certain probability, to receive all or none of the initial amount (i.e. the risky or gamble option). The sure option was formulated as either money retained from the initial amount (i.e. the gain frame) (e.g. 'Keep ¥ 20 out of a total of ¥ 50') or as money lost from the initial amount (i.e. the loss frame) (e.g. 'Lose ¥ 30 out of a total of ¥ 50'), presented in words. The gamble option was identical for both frames and was represented by a pie chart indicating the probability to receive all or none of the initial amount in the current trial. For both the gain frame and loss frame trials, the expected values of the two options in each trial were equivalent. For catch trials, the expected values of the sure option and the gamble option were extremely unbalanced (e.g. 'Keep ¥80 out of a total of ¥100' vs 'Keep all of the ¥100 with a probability of 40%'). These trials were introduced to allow us to examine whether a particular participant was actively engaged in the task. The behavioral test consisted of three sessions. Trial settings were the same for three sessions. Each session had 48 trials (16 different gain trials, 16 different loss trials and 16 different catch trials), ordered randomly. The payment procedure was conducted according to De Martino et al. (2006). The participants were informed that they were playing for real money at all times so their task was to be attending through the entire experiment which would allow them to maximize their final scores. At the end of the experiment they would receive a sum proportional (500:1) to what they earned during the experiment. See Gao et al. (2016) for more details about the behavioral test.

Two hundred and sixty-five participants were excluded from data analysis because of their low accuracy ($< 75\%$, mean accuracy = $57.9\% \pm 17.6\%$; other 1317 participants' accuracy = $90.5\% \pm 6.5\%$) in the catch condition. The excluded participants also showed much smaller framing effect ($4.44\% \pm 1.75\%$) compared with the remaining 1317 participants ($14.51\% \pm 0.34\%$), $t_{(1580)} = 9.16$, $P < 0.001$. Furthermore, we examined their response times when making choices (using a computer mouse to click on one of the two options presented on the screen). These participants responded faster when making decisions in both catch trials (mean = 1591 ms, SD = 551 ms) and experimental trials (mean = 1601 ms, SD = 544 ms) compared with the other participants (mean = 1776 ms, SD = 353 ms; mean = 1872 ms, SD = 376 ms), $t_{(1580)} = 6.639$, $P < 0.001$ for catch trials and $t_{(1580)} = 9.375$, $P < 0.001$ for experimental trials, indicating that these participants were careless when facing choices.

Genotyping

We collected ethylenediaminetetraacetic acid anti-coagulated venous blood samples from all the participants and then extracted genomic DNA from their whole blood using the QuickGene-610L Nucleic Acid Isolation System. The whole-genome genotyping was performed on Illumina Human OmniZhongHua-8 version 1 Chips using the standard Illumina genotyping protocol.

Gene selection and preprocessing

Twenty-six genes were selected based on the Kyoto Encyclopedia of Genes and Genomes database (KEGG; <http://www.genome.jp/kegg/>). We included dopamine genes involved in (i) dopamine synthesis [tyrosine hydroxylase (TH), aromatic-L-amino-acid decarboxylase gene (DDC), and vesicular monoamine transporter (VMAT2)], (ii) coding of dopamine receptors (DRD1–5), and (iii) dopamine transport and clearance [sodium-dependent dopamine transporter (DAT1, also named

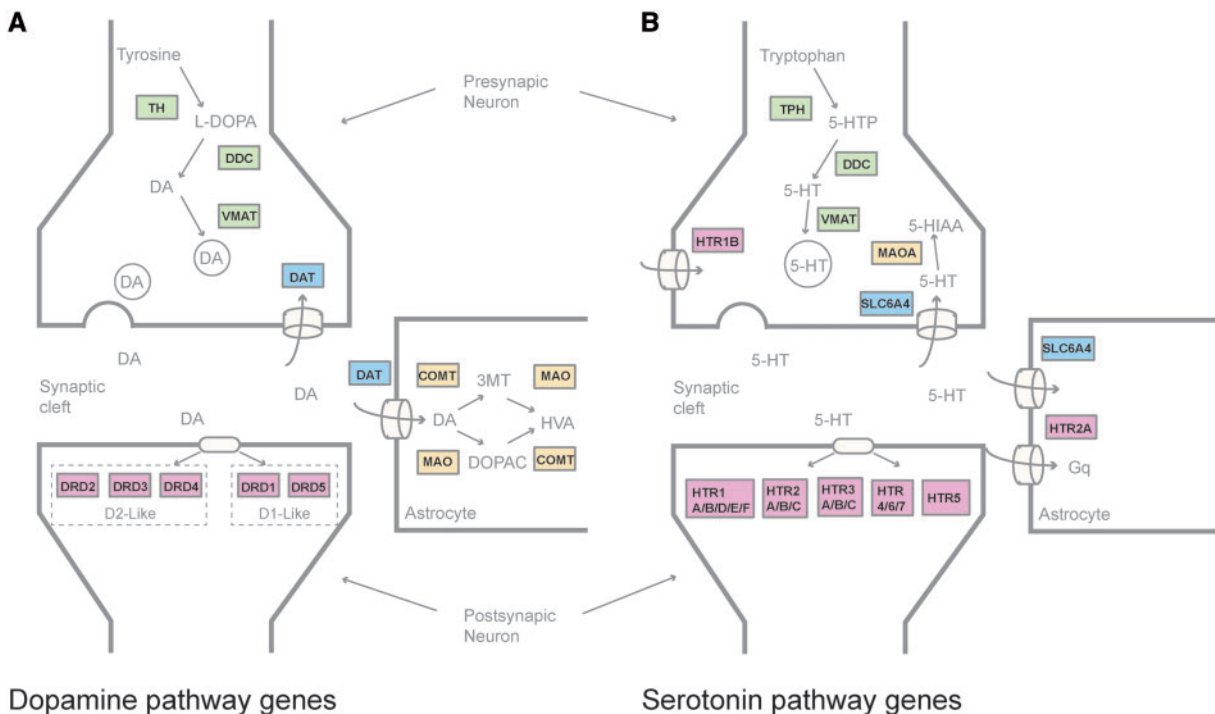


Fig. 1. Dopaminergic pathway genes and serotonergic pathway genes. Dopaminergic pathway genes (A) and serotonergic pathway genes (B) are represented in a stylized version of the synapse and include dopamine genes directly involved in synthesis (green), uptake (blue), and metabolism (yellow) and receptors (pink). Certain details, such as presynaptic auto-receptors, have been omitted for clarity. Adapted from the maps of dopaminergic synapse and serotonergic synapse in the Kyoto Encyclopedia of Genes and Genomes database (KEGG; <http://www.genome.jp/kegg/>); see also, Set et al., 2014.

SLC6A3), catechol-O-methyl transferase (COMT), amine oxidase A (MAOA) and amine oxidase B (MAOB)] (see also, Nemoda et al., 2011; Set et al., 2014) and serotonin genes involved in (i) serotonin biosynthesis [tryptophan 5-hydroxylase (TPH 1 and TPH2)], (ii) coding of serotonin receptors [5-hydroxytryptamine receptor (HTR1A/B/D/E/F, HTR2A/B/C, HTR3A/B/C/D/E, HTR4, HTR5A/B, HTR6-7, HTR1-4)], (iii) serotonin transport [sodium-dependent serotonin transporter (SLC6A4)] (see also Baou et al., 2016) (Figure 1). HTR3D and HTR3E genes were excluded from data analysis due to their lack of expression in the brain (Niesler et al., 2003; see also Bgee: Gene Expression Evolution, <http://bgee.unil.ch/>). DRD4, DRD5, HTR1A/B/D/F, HTR5B, HTRA2 and HTRA4 were also excluded from the final analysis due to the failure of extracting SNPs in the sample.

Preprocessing of GWA data was conducted in the following standard steps using PLINK (Purcell et al., 2007; Set et al., 2014): (i) we removed poorly genotyped SNPs, which were significantly depart from the HWE at a threshold of 10^{-4} or with minor allele frequency (MAF) below 0.1 or with genotyping rate below 0.05; (ii) we filtered out poor genotyped individuals with genotyping rate below 0.05 and (iii) we estimated population stratification and generated components of population stratification. To adjust for population stratification, two components indicating population stratification generated from the whole GWA data using the classical multidimensional scaling (MDS) method implemented in PLINK (Price et al., 2006; Purcell et al., 2007) were controlled in the following analysis. SNP extraction and filtering were conducted using PLINK (Purcell et al., 2007) and snpStats (Solé et al., 2006). For each gene, common SNPs were extracted according to hg19 coordinates.

Principle component analysis

All the following data analyses were conducted using R (The R Project for Statistical Computing, <http://www.r-project.org>). To avoid the collinearity arising from LD among SNPs of the same gene in the statistical model and reduce dimensionality of the genetic information, for each gene, we took all available SNPs in the GWA dataset and performed principal component analysis (PCA) on the SNPs within this gene to account for correlations due to LD (Wang and Abbott, 2008). Specifically, in the current study, each analyzed gene was represented by a set of eigenSNPs (PCs) accounting for at least 90% of the total variation of SNPs in this gene (Wang and Abbott, 2008; Set et al., 2014). The stronger the correlations between the SNP genotype scores (indicating the stronger the extent of LD among the SNPs), the fewer PCs are needed to capture the major variance in the original genotype scores. Consistent with densely spaced SNPs being in LD (Daly et al., 2001; Reich et al., 2001; Gabriel et al., 2002), SNPs within a gene were highly correlated (Table 1). For example, 6 eigenSNPs explained 90.2% of the variation of the DDC gene that contained 47 SNPs in our GWA dataset.

Gene-behavior association analysis

To test the joint effect of all variations in one gene on the susceptibility to framing, we employed a series of multiple partial-F tests with the susceptibility to framing (i.e. the rate of taking the gamble option in the loss frame minus this rate in the gain frame) as the dependent variable following the steps illustrated by previous studies (Wang and Abbott, 2008; Hibar et al., 2011a,b). Multiple partial-F tests are well suited for testing the effects of multiple predictors on a dependent variable. For each

Table 1. Summary of dopamine and serotonin genes and regression analysis

Pathway	Function	Gene	SNPs	PCs	%Var	R ² change	Adjusted R ² change	Partial-F	p _{unc}	p _{perm}	p _{emp}
Dopamine	Synthesis	TH	2	2	100	0.001	<0.001	0.712	0.491	0.484	0.485
		DDC	47	6	90	0.010	0.006	2.329	0.031*	0.031*	0.038*
		VMAT2	17	9	90	0.003	<0.001	0.501	0.875	0.878	0.862
	Transport/ Clearance	DAT1	16	6	91	0.005	<0.001	1.027	0.406	0.408	0.466
		COMT	18	6	91	0.012	0.009	2.648	0.015*	0.014*	0.027*
		MAOA	6	3	90	0.003	<0.001	1.143	0.331	0.325	0.346
	Receptor	MAOB	37	5	92	0.005	0.002	1.367	0.234	0.232	0.293
		DRD1	1	1	100	0.000	<0.001	0.097	0.756	0.756	0.780
		DRD2	16	8	90	0.004	<0.001	0.721	0.673	0.680	0.770
		DRD3	41	12	92	0.014	0.006	1.617	0.081	0.081	0.099
Serotonin	Synthesis	TPH1	2	2	100	0.001	<0.001	0.719	0.487	0.476	0.477
		TPH2	6	4	93	0.002	<0.001	0.519	0.721	0.718	0.753
	Transporter	SLC6A4	8	3	90	0.006	0.004	2.795	0.039*	0.038*	0.037*
	Receptor	HTR1E	16	6	91	0.007	0.003	1.545	0.160	0.158	0.199
		HTR2A	44	12	90	0.013	0.005	1.492	0.120	0.121	0.123
		HTR2B	3	2	100	0.001	<0.001	0.596	0.551	0.551	0.519
		HTR2C	22	8	90	0.006	<0.001	0.920	0.499	0.499	0.517
		HTR3A	4	4	100	0.001	<0.001	0.364	0.834	0.831	0.833
		HTR3B	22	6	90	0.001	<0.001	0.228	0.968	0.967	0.970
		HTR3C	2	1	99	0.000	<0.001	0.124	0.725	0.724	0.677
		HTR4	46	14	91	0.015	0.005	1.422	0.135	0.136	0.075
		HTR5A	7	4	92	0.006	0.003	1.866	0.114	0.114	0.118
		HTR6	2	1	100	0.000	<0.001	0.000	0.990	0.992	0.982
		HTR7	22	6	93	0.006	0.002	1.316	0.247	0.242	0.301
		HTRA1	34	9	91	0.007	<0.001	0.974	0.460	0.456	0.441
HTRA3	19	5	92	0.004	0.001	1.133	0.341	0.356	0.370		

PCs, the number of principal components; % Var, percentage of total variance captured by included PCs; p_{unc}, P value using multiple F-test; p_{perm}, permutation P value; p_{emp}, empirical P value.

*Means P < 0.05.

gene, the multiple partial-F test was conducted by firstly estimating the fit of a 'reduced model' of age, gender, and two components of population stratification (nuisance variables) on individuals' susceptibility to framing. Secondly, we estimated the fit of a second 'full model' with the nuisance variables and eigenSNPs of this gene (see the section *Principle component analysis*) on the same dependent variable. Each association test results in an F statistic, which indicates the joint effect of eigenSNPs of this gene on the behavior after controlling for the effects of age, gender and two components of population stratification. The multiple partial-F statistic was calculated for each gene using equation (1) (Hibar et al., 2011b). k is df(full) - df(reduced) and RSS is the residual sum of squares:

$$F_k, df(full) = \frac{RSS(reduced) - RSS(full)}{df(reduced) - df(full)} \cdot \frac{RSS(full)}{df(full)} \quad (1)$$

Of note, because the MAOA/B genes reside on the X-chromosome, females and males were analyzed separately to investigate the gene-behavior associations for these two genes.

Critically for our goal of identifying dopaminergic and serotonergic genes that are associated with the susceptibility to framing, variations across genes were essentially uncorrelated as shown by the very small proportion of variance explained by the other gene in the canonical correlation analysis (Weenink, 2003), mean variance explained by other gene = 0.62% ± 0.08% (SE) (see Table S1). Additionally, to examine the unique contribution of each gene to behavior while controlling for the contributions of other significant genes, we built a new regression model for each of the four genes that were identified to be

associated with framing effect (COMT, SLC6A, DDC and MAOB; see *Gene-behavior association results* for details). The new regression model included age, gender, two components of population stratification, as well as the eigenSNPs of the other three identified genes as nuisance variables. Controlling for the contributions of the other genes associated with the framing effect did not change the pattern of results (COMT: P = 0.028, SLC6A4: P = 0.038, DDC: P = 0.070 for all the participants, and MAOB: P = 0.029 for male participants), demonstrating the unique contribution of each gene to behavior.

Permutation tests

To guard against spurious associations and to further validate the above findings, we conducted the Monte Carlo permutation tests for each regression model (Hibar et al., 2011b; Set et al., 2014). This method is a widely accepted correction approach in statistical testing (Belmonte and Yurgelun-Todd, 2001; Nakagawa, 2004; Camargo et al., 2008; Gomez-Villegas et al., 2014), which resamples the total number of observations for certain times in order to estimate the regression coefficient in each shuffled sample and the probability of the estimated regression coefficients being greater than the observed regression coefficient (i.e. permutation P). This approach includes irregularities of the data in the estimation of the permutation probability (Cheverud, 2001).

Empirical tests

To guard against the possibility that the associations do not rise above the background association compared with the genome

at large, we compared *P* values in multiple partial-*F* tests of the genes on the dopaminergic and serotonergic pathways to comparable genes in the GWA dataset to generate an ‘empirical’ null distribution (Set *et al.*, 2014). Empirical *p* values were determined by comparing across the entire genome. A gene was considered comparable if (i) its SNPs generated the same number of principal components according to the procedure outlined above and (ii) it was represented by the same or similar number of SNPs. A range of SNPs was allowed to generate at least one hundred comparable genes, since an exact match produced too few comparable genes (see Supplementary Table S2). This typically occurred when there were a large number of SNPs within the gene.

Protein–protein interactions

Knowledge about a protein’s specific interaction map is an important prerequisite for a full understanding of its function. Here we used the STRING 10 (Search Tool for the Retrieval of Interacting Genes/Proteins) database (<http://string-db.org>, Szklarczyk *et al.*, 2015) to test the interactions between the proteins encoded by all the dopaminergic genes and serotonergic genes included in the current study. This database aims to provide a critical assessment and integration of protein–protein interactions, including direct (physical) as well as indirect (functional) associations, and generates an interaction confidence score for each interaction using four resources, including genomic context, high-throughput experiments, co-expression data, and previous studies.

Note, cellular functions are carried out by ‘modules’ made up of many species of interacting molecules (Hartwell *et al.*, 1999; Rives and Galitski, 2003). It is known that proteins of similar cellular functions tend to lie within a short distance in the interaction graph (Brun *et al.*, 2004). Thus, searching for interaction-modules may help us understand the relationship between the organization of a protein network and its function and thus provide independent evidence for the joint contribution of genes to a certain behavior. Using the ‘Clustering’ function implemented in STRING 10, we performed the MCL algorithm (inflation = 4), which is a widely used algorithm in clustering analysis (<http://www.micans.org/mcl/>, Brohee and Van Helden, 2006), to extract functional modules in our interaction graph (see Supplementary Figure S1).

SNP–SNP interactions

To estimate SNP–SNP interactions, we extended the eigenSNP approach by performing PCA on the set of regressors produced from a third-order interaction of the underlying SNP data. For example, if a gene contained three SNPs, we performed PCA on the set of seven regressors, resulting from three original SNPs, an additional three second-order interaction terms, and a further additional one third-order interaction term. Using the same procedure as outlined above, we took the set of eigenSNPs that explained at least 90% of the variance and included the concerning interaction terms in our computational model (see Supplementary Table S3).

Results

Behavioral results

Consistent with previous studies (De Martino *et al.*, 2006; Roiser *et al.*, 2009; Xu *et al.*, 2013; Gao *et al.*, 2016), a significant framing effect was observed for the rate of taking the risky or gamble

options: $59.75\% \pm 0.47\%$ (SE) in the loss frame vs $45.23\% \pm 0.46\%$ in gain the frame, $t_{(1316)} = 42.08$, $P < 0.0001$. The risk attitude change (i.e. the rate of taking the gamble option in the loss frame minus this rate in the gain frame) was defined as an individual’s susceptibility to framing in the following analysis. In line with previous studies (Fagley and Miller, 1990; Huang and Wang, 2010), a 2 (gender: Female vs Male) \times 2 (frame: gain vs loss) mixed measures analysis of variance (ANOVA) on the gambling rate revealed a significant interaction between gender and frame both before and after controlling for the potential effects of age, $F_{(1, 1315)} = 15.587$, $P < 0.001$, and $F_{(1, 1314)} = 14.701$, $P < 0.001$, with female participants evidencing a greater framing effect than male participants. In addition, when controlling for gender, linear regression analysis showed a marginally negative correlation between age and individuals’ susceptibility to framing, $\beta = -0.052$, $t = -1.903$, $P = 0.057$. This pattern was consistent with previous developmental studies (Mikels and Reed, 2009; Strough *et al.*, 2011). To exclude the effects of gender and age, these two factors were controlled as covariates in the analysis of gene–behavior association.

Gene–behavior association results

Consistent with our recent study showing the association between the COMT gene and the susceptibility to framing (Gao *et al.*, 2016), the regression analysis controlling for age, gender, and two principle components of population stratification indicated that eigenSNPs of the COMT gene explained 0.9% of the variance in individuals’ susceptibility to framing, adjusted R^2 change = 0.009, partial- $F = 2.648$, $P = 0.015$. Moreover, in line with Roiser *et al.* (2009) which demonstrated that 5-HTTLPR contributes to the individual differences, the entry of the eigenSNPs of the SLC6A4 gene accounted for 0.4% of the variance, adjusted R^2 change = 0.004, partial- $F = 2.795$, $P = 0.039$. Importantly, our results provide new evidence indicating that the DDC gene, which is involved in the synthesis of both serotonin and dopamine, was associated with individuals’ susceptibility to framing, adjusted R^2 change = 0.006, partial- $F = 2.329$, $P = 0.031$, accounting for 0.6% of the variance. None of the other genes were found to be predictive of the susceptibility to framing in decision-making in our sample (Table 1).

To guard against spurious associations, we conducted permutation tests for each regression model. After the Monte Carlo permutation test with 10 000 permutations of the behavioral data (individuals’ susceptibility to framing), the above results remained significant (Table 1), COMT: permutation $P = 0.014$, SLC6A4: permutation $P = 0.038$, DDC: permutation $P = 0.031$. Given that permutation test was conducted independently for each gene, it did not correct for multiple testing directly. Of note, the results obtained from the regression analyses did not survive the Bonferroni or false discovery rate (FDR) corrections for multiple testing (but see Discussion).

To test for the possibility that our evidence of association did not rise above the background association compared with the genome at large, we compared the fit of models between genes in the dopaminergic pathway and serotonergic pathway and other genes in the GWA dataset to generate an ‘empirical’ null distribution. Despite varying sizes of the comparison gene sets (see Supplementary Table S1), we found that, against the empirical null distributions, all the three genes showed significant differences (Table 1), COMT: empirical $P = 0.027$, SLC6A4: empirical $P = 0.037$, DDC: empirical $P = 0.038$.

Because the MAOA/B genes reside on the X-chromosome, there is substantial uncertainty regarding the interpretation of

allele scores across sex. Thus we estimated the model for these two genes in male and female participants separately. Results showed that the MAOB gene was associated with the susceptibility to framing in male participants, which accounted for 2.8% of the variation in male participants, adjusted R^2 change = 0.028, partial- F = 2.499, P = 0.031. The pattern remained the same in permutation test (P = 0.038) and empirical test (P = 0.043). This effect was absent in female participants. No effect was observed for the MAOA gene.

To address the question of whether there existed variations that could be explained by SNP-SNP interactions, we conducted PCA on regressors generated from first-order, second-order, and third-order interactions of SNPs within a gene. However, we found that incorporating SNP-SNP interactions did not improve model fittings of genes. The effects of the DDC gene and the SLC6A4 gene were similar to the results in single SNP analysis, while the effects of the COMT gene and the MAOB gene in single SNP analysis were now abolished. We did not find that the previously insignificant genes became significant after accounting for SNP-SNP interactions either (see Supplementary Table S3).

Using the STRING database to test the interactions between the proteins encoded by all genes included in our data analysis and extract functional modules in our interaction graph, we found that the four genes (COMT, SLC6A4, DDC and MAOB) associated with the susceptibility to framing in the current study were shown to interact with each other and were clustered into the same module (see Supplementary Figure S1).

Given the PCA analysis here did not allow us to identify which SNPs constituted the principal components that contributed to the framing effect (Harris, 1975), we examined directly how identified gene-behavior associations were distributed across SNPs in the identified genes. SNPs associated with individual susceptibility to framing are shown in Figure S2 (Supplementary data). For the COMT gene, although rs4680 did not significantly contribute to the framing effect, a SNP in LD with rs4680, rs165656 (1000Genomes, phase_3, Han Chinese in Beijing, China (CHB): r^2 = 0.934, D' = 1.000; 1000Genomes, phase_3, Southern Han Chinese, China (CHS): r^2 = 0.816, D' = 0.948; see also Zhang et al., 2007), was significantly associated with the susceptibility to framing (adjusted R^2 change = 0.004, partial- F = 5.842, P = 0.016). See Supplementary data for more details about individual SNP associations.

Discussion

Twin studies yield a moderate heritability of the susceptibility to framing in decision-making (Simonson and Sela, 2011; Cesarini et al., 2012; Cronqvist and Siegel, 2012). Other studies also demonstrated important roles of genetic variations on dopaminergic and serotonergic pathways in decision-making under risks (Crısan et al., 2009; Dreber et al., 2009; Kuhnen and Chiao, 2009; He et al., 2010; Frydman et al., 2011; Heitland et al., 2012; Reuter et al., 2013; Set et al., 2014). By extending the few studies on the genetic basis of the framing effect (Crısan et al., 2009; Roiser et al., 2009; Gao et al., 2016) and by using a gene-based PCReg approach, we investigated directly the relationship between 26 genes within the dopaminergic and the serotonergic pathways and individuals' susceptibility to framing in decision-making.

Replicating the previous SNP-based studies, which had relatively small sample size, our results confirmed that genetic variations in the COMT gene (Gao et al., 2016) and the SLC6A4 gene (Crısan et al., 2009; Roiser et al., 2009) contribute to the individual differences in susceptibility to framing. Although COMT rs4680

did not significantly contribute to the framing effect in the current sample, a SNP in LD with rs4680, rs165656, was significantly associated with the susceptibility to framing. This result raised the possibility that the previously identified effect of rs4680 (Gao et al., 2016) might be due to its LD with rs165656. Moreover, the three previous studies merely investigated a single polymorphism in the candidate gene; an important advance the current study made was that we used the gene-based PCReg approach to search for genes associated with susceptibility to framing in a larger sample. This approach takes into account common variations within a candidate gene to avoid the influence of the LD among SNPs and is more efficient than the SNP-based association method (Wang and Abbott, 2008; Wang et al., 2010; Set et al., 2014). Compared with methods that rely on data from SNPs independently, the gene-based approach is less susceptible to erroneous findings due to genetic differences between populations (Neale and Sham, 2004; Schaid, 2004). Based on this gene-based approach, our results identified a new gene, DDC, for the individual differences in susceptibility to framing.

In humans, the DDC gene encodes DDC, an enzyme that catalyzes the decarboxylation of 5-hydroxytryptophan to serotonin and L-DOPA to dopamine (Sumi-Ichinose et al., 1992). Given that this gene is involved in both the dopaminergic pathway and the serotonergic pathway, two pathways that play critical roles in emotional processing (Salgado-Pineda et al., 2005; Alcaro et al., 2007; Merens et al., 2007; Cools et al., 2008; Harmer, 2008; Badgaiyan et al., 2009), this gene may contribute to a series of processes involved in affective processing. For example, SNP-based association studies demonstrated that the DDC gene is associated with emotional dysregulation such as anxiety (Costas et al., 2010) and bipolar disorder (Børghlum et al., 2003). In line with this suggestion, our results provide new evidence that the genetic variations of the DDC gene contribute to individual differences in susceptibility to framing, which is related to negative emotional processing in decision-making (De Martino et al., 2006, 2008; Druckman and McDermott, 2008; Roiser et al., 2009; Miu and Crısan, 2011; Xu et al., 2013; Gao et al., 2016). These findings indicate that genetic variations in the DDC gene may be a candidate gene for genetic studies on other affective decision-making (e.g. Iowa gambling task).

The effects of genes are not expressed directly at the level of behavior, but are mediated by their effects on brain regions responsible for specific cognitive and emotional processes (Bigos and Weinberger, 2010). A pathway-based approach allows us to utilize prior knowledge regarding relationships between genes and the brain and to shed light on the neural mechanisms underlying the gene-behavior association. Neuroimaging and lesion studies have shown that the prefrontal (PFC)-amygdala circuitry plays important role in the framing effect (De Martino et al., 2006, 2010; Roiser et al., 2009; Gao et al., 2016). In line with this evidence, all the genes associated with the susceptibility to framing in the current study showed high expression in these circuits. Specifically, while the COMT gene and the MAOB gene are important determinants of dopamine flux in the PFC (Nemoda et al., 2011), the SLC6A4 expression modulates amygdala activation (Munafò et al., 2008; Kobiella et al., 2011). Moreover, the expression of the DDC gene has been observed in the PFC and limbic areas in humans (Gjedde et al., 1991; Sumi-Ichinose et al., 1995; Ikemoto et al., 1999) and other primates (Kitahama et al., 1988). Given the genetic imaging studies demonstrating that both the SLC6A4 gene 5-HTTLPR variation (Roiser et al., 2009) and the COMT Val158Met polymorphism (Gao et al., 2016) influence the susceptibility to framing via the PFC-

amygdala coupling, it is conceivable that genetic variations in the *DDC* gene and the *MAOB* gene may modulate the individual differences in susceptibility to framing via their impacts on the PFC-amygdala circuitry. This hypothesis needs to be tested by further genetic imaging studies.

The current study raises a few more implications for future research. First, in line with a previous study suggesting that the *MAOB* polymorphisms are related to negative emotionality personality (Dlugos et al., 2009), our results provide new evidence linking genetic variations of *MAOB* to the susceptibility to framing, which is associated with negative emotional processing. However, this effect exists in male participants, but not in female participants. This sex difference is consistent with previous neuropsychological studies demonstrating that the platelet MAO activity is associated with sex-differentiated features in cognitive processing (Klinterberg et al., 1987; Tadić et al., 2007) and SNP-based association studies reporting sex-specific effects of the *MAOB* gene variations on different phenotypes [e.g. Parkinson's disease (Kelada et al., 2002; Kang et al., 2006)]. However, given that the number of male participants was relatively small in the current study, future studies are needed to examine the relationship between the *MAOB* gene and the framing effect. Second, using the protein-protein interaction information and clustering analysis in STRING database, we found that proteins encoded by the four genes associated with the framing effect in the current study are also clustered into the same functional module, demonstrating their strong interactions with each other (Szkłarczyk et al., 2015). Thus, whether and how the interactions between these four genes influence the individual differences in susceptibility to framing is an important question that remains to be investigated. Third, consistent with the notion that most common genetic variants individually or in combination explain only a small proportion of heritability using GWA method (Manolio et al., 2009; Zuk et al., 2014), each of the three genes identified here contributed to about 0.5%–1.0% variance of individuals' susceptibility to framing, which was only a small proportion of the heritability estimated in twin studies (Simonson and Sela, 2011; Cesarini et al., 2012; Cronqvist and Siegel, 2012). This implies that genetic variations of other underestimated pathways, especially pathways related to affective processing (e.g. the oxytocin pathway and the vasopressin pathway, cf. Ebstein et al., 2012; Neumann and Landgraf, 2012), might further explain the remaining variance of individuals' susceptibility to framing. Under-reported genetic variants of smaller effect, rare variants that are poorly detected by available genotyping arrays, and low power to detect gene-gene interactions, may also contribute to this missing heritability (Manolio et al., 2009).

Although the current study adds to the limited information on the genetic basis of individual differences in susceptibility to framing, it should be noticed that for the exploratory aim of the study, we examined 26 genes, which may create inflation of type I error rate. Moreover, the fact that none of the identified associations survived Bonferroni or FDR corrections for multiple testing may raise the concern that the significant associations observed could emerge simply by chance. However, several factors mitigate this concern. First, replicating the results of previous SNP-based studies (Roiser et al., 2009; Gao et al., 2016), the *COMT* gene and the *SLC6A4* gene were associated with the framing effect in the current study. This replication may demonstrate the validity of the current study. Second, in support of our conclusions, proteins encoded by the four genes identified here are clustered into the same functional module in the independent protein-protein interaction analysis, suggesting that

these four proteins are crucial components of similar cellular processes (Hartwell et al., 1999; Rives and Galitski, 2003; Brun et al., 2004), which might in turn influence the same or similar behaviors (i.e. the framing effect). Third, all our results passed the empirical tests that aimed to guard against the possibility that the observed associations do not rise above the background association compared with the genome at large (Set et al., 2014). Finally, although multiple corrections such as Bonferroni or FDR correction reduce type I errors, stringent corrections may lead to enlarged type II errors (Nakagawa, 2004). One of the better ways to control for type I errors is to examine these results in another larger sample. Future study investigating the association between *DDC* gene and the susceptibility to framing will be essential.

In conclusion, the current study replicated previous SNP-based association studies by demonstrating that genetic variations of the *SLC6A4* gene and the *COMT* gene contribute to the susceptibility to framing during decision-making. More importantly, the current study provides the first evidence for the role of the *DDC* gene (and, to a less extent, the *MAOB* gene) in individual susceptibility to framing. These findings shed light on our understanding of the genetic basis underlying individual differences in decision-making.

Supplementary data

Supplementary data are available at SCAN online.

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Reference

- Alcaro, A., Huber, R., Panksepp, J. (2007). Behavioral functions of the mesolimbic dopaminergic system: an affective neuroethological perspective. *Brain Research Reviews*, *56*(2), 283–321.
- Badgaiyan, R.D., Fischman, A.J., Alpert, N.M. (2009). Dopamine release during human emotional processing. *Neuroimage*, *47*(4), 2041–5.
- Baou, M., Boumba, V.A., Petrikis, P., Rallis, G., Vougiouklakis, T., Mavreas, V. (2016). A review of genetic alterations in the serotonin pathway and their correlation with psychotic diseases and response to atypical antipsychotics. *Schizophrenia Research*, *170*(1), 18–29.
- Bellivier, F., Henry, C., Szöke, A., et al. (1998). Serotonin transporter gene polymorphisms in patients with unipolar or bipolar depression. *Neuroscience Letters*, *255*(3), 143–6.
- Belmonte, M., Yurgelun-Todd, D. (2001). Permutation testing made practical for functional magnetic resonance image analysis. *IEEE Transactions on Medical Imaging*, *20*(3), 243–8.
- Bevilacqua, L., Goldman, D. (2011). Genetics of emotion. *Trends in Cognitive Sciences*, *15*(9), 401–8.

- Bigos, K.L., Weinberger, D.R. (2010). Imaging genetics—days of future past. *Neuroimage*, *53*(3), 804–9.
- Børghlum, A., Kirov, G., Craddock, N., et al. (2003). Possible parent-of-origin effect of DOPA decarboxylase in susceptibility to bipolar affective disorder. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, *117*(1), 18–22.
- Brohee, S., Van Helden, J. (2006). Evaluation of clustering algorithms for protein–protein interaction networks. *BMC Bioinformatics*, *7*(1), 1.
- Brun, C., Herrmann, C., Guénoche, A. (2004). Clustering proteins from interaction networks for the prediction of cellular functions. *BMC Bioinformatics*, *5*(1), 1.
- Camargo, A., Azuaje, F., Wang, H., Zheng, H. (2008). Permutation-based statistical tests for multiple hypotheses. *Source Code for Biology and Medicine*, *3*, 15.
- Canli, T., Omura, K., Haas, B.W., Fallgatter, A., Constable, R.T., Lesch, K.P. (2005). Beyond affect: a role for genetic variation of the serotonin transporter in neural activation during a cognitive attention task. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(34), 12224–9.
- Cesarini, D., Johannesson, M., Magnusson, P.K., Wallace, B. (2012). The behavioral genetics of behavioral anomalies. *Management Science*, *58*(1), 21–34.
- Cheverud, J.M. (2001). A simple correction for multiple comparisons in interval mapping genome scans. *Heredity (Edinb)*, *87*(Pt 1), 52–8.
- Cools, R., Roberts, A.C., Robbins, T.W. (2008). Serotonergic regulation of emotional and behavioural control processes. *Trends in Cognitive Sciences*, *12*(1), 31–40.
- Costas, J., Gratacos, M., Escaramis, G., et al. (2010). Association study of 44 candidate genes with depressive and anxiety symptoms in post-partum women. *Journal of Psychiatric Research*, *44*(11), 717–24.
- Cronqvist, H., Siegel, S. (2012). Why do individuals exhibit investment biases. *Claremont McKenna College Robert Day School of Economics and Finance Research Paper (2012-01)*.
- Crişan, L.G., Pană, S., Vulturar, R., et al. (2009). Genetic contributions of the serotonin transporter to social learning of fear and economic decision making. *Social Cognitive and Affective Neuroscience*, *4*(4), 399–408.
- Daly, M.J., Rioux, J.D., Schaffner, S.F., Hudson, T.J., Lander, E.S. (2001). High-resolution haplotype structure in the human genome. *Nature Genetics*, *29*(2), 229–32.
- De Martino, B., Camerer, C.F., Adolphs, R. (2010). Amygdala damage eliminates monetary loss aversion. *Proceedings of the National Academy of Sciences*, *107*(8), 3788–92.
- De Martino, B., Harrison, N.A., Knafo, S., Bird, G., Dolan, R.J. (2008). Explaining enhanced logical consistency during decision making in autism. *Journal of Neuroscience*, *28*(42), 10746–50.
- De Martino, B., Kumaran, D., Seymour, B., Dolan, R.J. (2006). Frames, biases, and rational decision-making in the human brain. *Science*, *313*(5787), 684–7.
- Dlugos, A.M., Palmer, A.A., de Wit, H. (2009). Negative emotionality: monoamine oxidase B gene variants modulate personality traits in healthy humans. *Journal of Neural Transmission*, *116*(10), 1323–34.
- Drabant, E.M., Hariri, A.R., Meyer-Lindenberg, A., et al. (2006). Catechol O-methyltransferase val158met genotype and neural mechanisms related to affective arousal and regulation. *Archives of General Psychiatry*, *63*(12), 1396–406.
- Dreber, A., Apicella, C.L., Eisenberg, D.T., et al. (2009). The 7R polymorphism in the dopamine receptor D 4 gene (DRD4) is associated with financial risk taking in men. *Evolution and Human Behavior*, *30*(2), 85–92.
- Druckman, J.N. (2004). Political preference formation: competition, deliberation, and the (ir) relevance of framing effects. *American Political Science Review*, *98*(04), 671–86.
- Druckman, J.N., McDermott, R. (2008). Emotion and the framing of risky choice. *Political Behavior*, *30*(3), 297–321.
- Ebstein, R.P., Knafo, A., Mankuta, D., Chew, S.H., San Lai, P. (2012). The contributions of oxytocin and vasopressin pathway genes to human behavior. *Hormones and Behavior*, *61*(3), 359–79.
- Enoch, M.A., Xu, K., Ferro, E., Harris, C.R., Goldman, D. (2003). Genetic origins of anxiety in women: a role for a functional catechol-O-methyltransferase polymorphism. *Psychiatric Genetics*, *13*(1), 33–41.
- Fabbri, C., Marsano, A., Serretti, A. (2013). Genetics of serotonin receptors and depression: state of the art. *Current Drug Targets*, *14*(5), 531–48.
- Fagley, N.S., Miller, P.M. (1990). The effect of framing on choice interactions with risk-taking propensity, cognitive style, and sex. *Personality and Social Psychology Bulletin*, *16*(3), 496–510.
- Frydman, C., Camerer, C., Bossaerts, P., Rangel, A. (2011). MAOA-L carriers are better at making optimal financial decisions under risk. *Proceedings of the Royal Society of London B: Biological Sciences*, *278*(1714), 2053–9.
- Gabriel, S.B., Schaffner, S.F., Nguyen, H., et al. (2002). The structure of haplotype blocks in the human genome. *Science*, *296*(5576), 2225–9.
- Gadow, K.D., Pinsonneault, J.K., Perlman, G., Sadee, W. (2014). Association of dopamine gene variants, emotion dysregulation and ADHD in autism spectrum disorder. *Research in Developmental Disabilities*, *35*(7), 1658–65.
- Gao, X., Gong, P., Liu, J., et al. (2016). COMT Val158Met polymorphism influences the susceptibility to framing in decision-making: OFC-amygdala functional connectivity as a mediator. *Human Brain Mapping*, *37*(5), 1880–92.
- Gjedde, A., Reith, J., Dyve, S., et al. (1991). Dopa decarboxylase activity of the living human brain. *Proceedings of the National Academy of Sciences*, *88*(7), 2721–5.
- Gomez-Villegas, M.A., Salazar, I., Sanz, L. (2014). A Bayesian decision procedure for testing multiple hypotheses in DNA microarray experiments. *Statistical Applications in Genetics and Molecular Biology*, *13*(1), 49–65.
- Hariri, A.R., Mattay, V.S., Tessitore, A., et al. (2002). Serotonin transporter genetic variation and the response of the human amygdala. *Science*, *297*(5580), 400–3.
- Harmer, C.J. (2008). Serotonin and emotional processing: does it help explain antidepressant drug action? *Neuropharmacology*, *55*(6), 1023–8.
- Harris, R. (1975). *A Primer of Multivariate Statistics*. New York, NY: Academic Press.
- Hartwell, L.H., Hopfield, J.J., Leibler, S., Murray, A.W. (1999). From molecular to modular cell biology. *Nature*, *402*, C47–52.
- He, Q., Xue, G., Chen, C., et al. (2010). Serotonin transporter gene-linked polymorphic region (5-HTTLPR) influences decision making under ambiguity and risk in a large Chinese sample. *Neuropharmacology*, *59*(6), 518–26.
- Heinz, A., Braus, D.F., Smolka, M.N., et al. (2005). Amygdala-prefrontal coupling depends on a genetic variation of the serotonin transporter. *Nature Neuroscience*, *8*(1), 20–1.
- Heinz, A., Smolka, M.N. (2006). The effects of catechol O-methyltransferase genotype on brain activation elicited by affective stimuli and cognitive tasks. *Reviews in the Neurosciences*, *17*(3), 359–67.
- Heitland, I., Oosting, R., Baas, J., Massar, S., Kenemans, J., Böcker, K. (2012). Genetic polymorphisms of the dopamine and

- serotonin systems modulate the neurophysiological response to feedback and risk taking in healthy humans. *Cognitive, Affective, & Behavioral Neuroscience*, **12**(4), 678–91.
- Hibar, D.P., Kohannim, O., Stein, J.L., Chiang, M.-C., Thompson, P.M. (2011a). Multilocus genetic analysis of brain images. *Frontiers in Genetics*, **2**, 73.
- Hibar, D.P., Stein, J.L., Kohannim, O., et al. (2011b). Voxelwise gene-wide association study (vGeneWAS): multivariate gene-based association testing in 731 elderly subjects. *Neuroimage*, **56**(4), 1875–91.
- Hill, E., Berthoz, S., Frith, U. (2004). Brief report: cognitive processing of own emotions in individuals with autistic spectrum disorder and in their relatives. *J Autism and Developmental Disorders*, **34**(2), 229–35.
- Huang, Y., Wang, L. (2010). Sex differences in framing effects across task domain. *Personality and Individual Differences*, **48**(5), 649–53.
- Ikemoto, K., Kitahama, K., Nishimura, A., et al. (1999). Tyrosine hydroxylase and aromatic l-amino acid decarboxylase do not coexist in neurons in the human anterior cingulate cortex. *Neuroscience Letters*, **269**(1), 37–40.
- Jonassen, R., Landrø, N. (2014). Serotonin transporter polymorphisms (5-HTTLPR) in emotion processing: implications from current. *Neurobiology. Progress in Neurobiology*, **117**, 41–53.
- Kahneman, D., Tversky, A. (1979). Prospect theory: an analysis of decision under risk. *Econometrica: Journal of the Econometric Society*, 263–91.
- Kahneman, D., Tversky, A. (1984). Choices, values, and frames. *American Psychologist*, **39**(4), 341.
- Kang, S.J., Scott, W.K., Li, Y.J., et al. (2006). Family-based case-control study of MAOA and MAOB polymorphisms in Parkinson disease. *Movement Disorders*, **21**(12), 2175–80.
- Kelada, S.N., Costa-Mallen, P., Costa, L.G., et al. (2002). Gender difference in the interaction of smoking and monoamine oxidase B intron 13 genotype in Parkinson's disease. *Neurotoxicology*, **23**(4), 515–9.
- Kersting, A., Kroker, K., Horstmann, J., et al. (2007). Association of MAO-A variant with complicated grief in major depression. *Neuropsychobiology*, **56**(56), 191–6.
- Kia-Keating, B.M., Glatt, S.J., Tsuang, M.T. (2007). Meta-analyses suggest association between COMT, but not HTR1B, alleles, and suicidal behavior. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, **144B**(8), 1048–53.
- Kitahama, K., Denoyer, M., Raynaud, B., et al. (1988). Immunohistochemistry of aromatic l-amino acid decarboxylase in the cat forebrain. *Journal of Comparative Neurology*, **270**(3), 337–53.
- Klei, L., Luca, D., Devlin, B., Roeder, K. (2008). Pleiotropy and principal components of heritability combine to increase power for association analysis. *Genetic Epidemiology*, **32**(1), 9–19.
- Klinteberg, B., Levander, S.E., Orelund, L., Asberg, M., Schalling, D. (1987). Neuropsychological correlates of platelet monoamine oxidase (MAO). Activity in female and male subjects. *Biological Psychology*, **24**(3), 237–52.
- Kobiella, A., Reimold, M., Ulshöfer, D., et al. (2011). How the serotonin transporter 5-HTTLPR polymorphism influences amygdala function: the roles of in vivo serotonin transporter expression and amygdala structure. *Translational Psychiatry*, **1**(8), e37.
- Kuhnen, C.M., Chiao, J.Y. (2009). Genetic determinants of financial risk taking. *PLoS ONE*, **4**(2), e4362.
- Kuhberger, A., Schulte-Mecklenbeck, M., Perner, J. (1999). The effects of framing, reflection, probability, and payoff on risk preference in choice tasks. *Organizational Behavior and Human Decision Processes*, **78**(3), 204–31.
- Lasky-Su, J.A., Faraone, S.V., Glatt, S.J., Tsuang, M.T. (2005). Meta-analysis of the association between two polymorphisms in the serotonin transporter gene and affective disorders. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, **133**(1), 110–5.
- Lotrich, F.E., Pollock, B.G. (2004). Meta-analysis of serotonin transporter polymorphisms and affective disorders. *Psychiatric Genetics*, **14**(3), 121–9.
- Ma, Y., Li, B., Wang, C., et al. (2014). 5-HTTLPR polymorphism modulates neural mechanisms of negative self-reflection. *Cerebral Cortex*, **24**(9), 2421–9.
- Manolio, T.A., Collins, F.S., Cox, N.J., et al. (2009). Finding the missing heritability of complex diseases. *Nature*, **461**(7265), 747–53.
- McGrath, M., Kawachi, I., Ascherio, A., Colditz, G.A., Hunter, D.J., De Vivo, I. (2004). Association between catechol-O-methyltransferase and phobic anxiety. *American Journal of Psychiatry*, **161**(9), 1703–5.
- McNeil, B.J., Pauker, S.G., Sox, H.C., Jr, Tversky, A. (1982). On the elicitation of preferences for alternative therapies. *The New England Journal of Medicine*, **306**(21), 1259–62.
- Merens, W., Willem Van der Does, A.J., Spinhoven, P. (2007). The effects of serotonin manipulations on emotional information processing and mood. *Journal of Affective Disorders*, **103**(1–3), 43–62.
- Meyer-Lindenberg, A. (2010). Imaging genetics of schizophrenia. *Dialogues in Clinical Neuroscience*, **12**(4), 449–56.
- Mikels, J.A., Reed, A.E. (2009). Monetary losses do not loom large in later life: Age differences in the framing effect. *The Journals of Gerontology Series B: Psychological Sciences and Social Sciences*, **64**(4), 457–60.
- Miu, A.C., Crişan, L.G. (2011). Cognitive reappraisal reduces the susceptibility to the framing effect in economic decision making. *Personality and Individual Differences*, **51**(4), 478–82.
- Montag, C., Buckholtz, J.W., Hartmann, P., et al. (2008). COMT genetic variation affects fear processing: psychophysiological evidence. *Behavioral Neuroscience*, **122**(4), 901–9.
- Munafò, M.R., Brown, S.M., Hariri, A.R. (2008). Serotonin transporter (5-HTTLPR) genotype and amygdala activation: a meta-analysis. *Biological Psychiatry*, **63**(9), 852–7.
- Munafò, M.R., Durrant, C., Lewis, G., Flint, J. (2009). Gene × environment interactions at the serotonin transporter locus. *Biological Psychiatry*, **65**(3), 211–9.
- Nakagawa, S. (2004). A farewell to Bonferroni: the problems of low statistical power and publication bias. *Behavioral Ecology*, **15**(6), 1044–5.
- Neale, B.M., Sham, P.C. (2004). The future of association studies: gene-based analysis and replication. *American Journal of Human Genetics*, **75**(3), 353–62.
- Nemoda, Z., Szekely, A., Sasvari-Szekely, M. (2011a). Psychopathological aspects of dopaminergic gene polymorphisms in adolescence and young adulthood. *Neuroscience & Biobehavioral Reviews*, **35**(8), 1665–86.
- Neumann, I.D., Landgraf, R. (2012). Balance of brain oxytocin and vasopressin: implications for anxiety, depression, and social behaviors. *Trends in Neurosciences*, **35**(11), 649–59.
- Niesler, B., Frank, B., Kapeller, J., Rappold, G.A. (2003). Cloning, physical mapping and expression analysis of the human 5-HT₃ serotonin receptor-like genes HTR3C, HTR3D and HTR3E. *Gene*, **310**, 101–11.
- Ohara, K., Nagai, M., Suzuki, Y., Ohara, K. (1998). Low activity allele of catechol-o-methyltransferase gene and Japanese unipolar depression. *Neuroreport*, **9**(7), 1305–8.

- Olsson, C.A., Byrnes, G.B., Anney, R.J., et al. (2007). COMT Val(158)Met and 5HTTLPR functional loci interact to predict persistence of anxiety across adolescence: results from the Victorian Adolescent Health Cohort Study. *Genes, Brain and Behavior*, *6*(7), 647–52.
- Opmeer, E.M., Kortekaas, R., Aleman, A. (2010). Depression and the role of genes involved in dopamine metabolism and signalling. *Progress in Neurobiology*, *92*(2), 112–33.
- Price, A.L., Patterson, N.J., Plenge, R.M., Weinblatt, M.E., Shadick, N.A., Reich, D. (2006). Principal components analysis corrects for stratification in genome-wide association studies. *Nature Genetics*, *38*(8), 904–9.
- Purcell, S., Neale, B., Todd-Brown, K., et al. (2007). PLINK: a tool set for whole-genome association and population-based linkage analyses. *The American Journal of Human Genetics*, *81*(3), 559–75.
- Reich, D.E., Cargill, M., Bolk, S., et al. (2001). Linkage disequilibrium in the human genome. *Nature*, *411*(6834), 199–204.
- Reuter, M., Felten, A., Penz, S., Mainzer, A., Markett, S., Montag, C. (2013). The influence of dopaminergic gene variants on decision making in the ultimatum game. *Frontiers in Human Neuroscience*, *7*, 242.
- Rives, A.W., Galitski, T. (2003). Modular organization of cellular networks. *Proceedings of the National Academy of Sciences*, *100*(3), 1128–33.
- Roiser, J.P., de Martino, B., Tan, G.C., et al. (2009). A genetically mediated bias in decision making driven by failure of amygdala control. *Journal of Neuroscience*, *29*(18), 5985–91.
- Roohi, J., Devincenzi, C.J., Hatchwell, E., Gadow, K.D. (2009). Association of a monoamine oxidase-A gene promoter polymorphism with ADHD and anxiety in boys with autism spectrum disorder. *Journal of Autism & Developmental Disorders*, *39*(1), 67–74.
- Salgado-Pineda, P., Delaveau, P., Blin, O., Nieoullon, A. (2005). Dopaminergic contribution to the regulation of emotional perception. *Clinical Neuropharmacology*, *28*(5), 228–37.
- Schadt, E.E. (2009). Molecular networks as sensors and drivers of common human diseases. *Nature*, *461*(7261), 218–23.
- Schaid, D.J. (2004). Evaluating associations of haplotypes with traits. *Genetic Epidemiology*, *27*(4), 348–64.
- Scharinger, C., Rabl, U., Sitte, H.H., Pezawas, L. (2010). Imaging genetics of mood disorders. *Neuroimage*, *53*(3), 810–21.
- Schupp, H.T., Junghofer, M., Weike, A.I., Hamm, A.O. (2003). Emotional facilitation of sensory processing in the visual cortex. *Psychological Science*, *14*(1), 7–13.
- Set, E., Saez, I., Zhu, L., et al. (2014). Dissociable contribution of prefrontal and striatal dopaminergic genes to learning in economic games. *Proceedings of the National Academy of Sciences*, *111*(26), 9615–20.
- Sharp, D.J., Salter, S.B. (1997). Project escalation and sunk costs: a test of the international generalizability of agency and prospect theories. *Journal of International Business Studies*, 101–21.
- Simonson, I., Sela, A. (2011). On the heritability of consumer decision making: an exploratory approach for studying genetic effects on judgment and choice. *Journal of Consumer Research*, *37*(6), 951–66.
- Smolka, M.N., Schumann, G., Wrase, J., et al. (2005). Catechol-O-methyltransferase val158met genotype affects processing of emotional stimuli in the amygdala and prefrontal cortex. *Journal of Neuroscience*, *25*(4), 836–42.
- Solé, X., Guinó, E., Valls, J., Iniesta, R., Moreno, V. (2006). SNPStats: a web tool for the analysis of association studies. *Bioinformatics*, *22*(15), 1928–9.
- Strough, J., Karns, T.E., Schlosnagle, L. (2011). Decision-making heuristics and biases across the life span. *Annals of the New York Academy of Sciences*, *1235*(1), 57–74.
- Sumi-Ichinose, C., Ichinose, H., Takahashi, E., Hori, T., Nagatsu, T. (1992). Molecular cloning of genomic DNA and chromosomal assignment of the gene for human aromatic l-amino acid decarboxylase, the enzyme for catecholamine and serotonin biosynthesis. *Biochemistry*, *31*(8), 2229–38.
- Sumi-Ichinose, C., Hasegawa, S., Ichinose, H., et al. (1995). Analysis of the alternative promoters that regulate tissue-specific expression of human aromatic l-amino acid decarboxylase. *Journal of Neurochemistry*, *64*(2), 514–24.
- Szklarczyk, D., Franceschini, A., Wyder, S., et al. (2015). STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Research* *43*(Database issue), D447–52.
- Tadić, A., Rujescu, D., Müller, M.J., et al. (2007). A monoamine oxidase B gene variant and short-term antidepressant treatment response. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, *31*(7), 1370–7.
- Tversky, A., Kahneman, D. (1981). The framing of decisions and the psychology of choice. *Science* *211*(4481), 453–8.
- Uher, R., McGuffin, P. (2010). The moderation by the serotonin transporter gene of environmental adversity in the etiology of depression: 2009 update. *Molecular Psychiatry*, *15*(1), 18–22.
- Wang, K., Abbott, D. (2008). A principal components regression approach to multilocus genetic association studies. *Genetic Epidemiology*, *32*(2), 108–18.
- Wang, K., Li, M., Bucan, M. (2007). Pathway-based approaches for analysis of genomewide association studies. *The American Journal of Human Genetics*, *81*(6), 1278–83.
- Wang, K., Li, M., Hakonarson, H. (2010). Analysing biological pathways in genome-wide association studies. *Nature Reviews Genetics*, *11*(12), 843–54.
- Wang, X., Wang, X., Ma, H. (1999). *Manual of Mental Health Rating Scales*. Beijing: Chinese Mental Health Journal Publisher.
- Weenink, D. (2003). Canonical correlation analysis. *Proceedings of the Institute of Phonetic Sciences of the University of Amsterdam*, *25*, 81–99.
- Williams, L.M., Gatt, J.M., Grieve, S.M., et al. (2010). COMT Val(108/158)Met polymorphism effects on emotional brain function and negativity bias. *Neuroimage*, *53*(3), 918–25.
- Xu, P., Gu, R., Broster, L.S., et al. (2013). Neural basis of emotional decision making in trait anxiety. *Journal of Neuroscience*, *33*(47), 18641–53.
- Zhang, K., Gao, J., An, C., et al. (2007). An association study between catechol-O-methyltransferase gene and mental retardation in the Chinese Han population. *Neuroscience Letters*, *419*(1), 83–7.
- Zuk, O., Schaffner, S.F., Samocha, K., et al. (2014). Searching for missing heritability: designing rare variant association studies. *Proceedings of the National Academy of Sciences*, *111*(4), E455–64.
- Zung, W.W. (1965). A self-rating depression scale. *Archives of General Psychiatry*, *12*(1), 63–70.
- Zung, W.W. (1971). A rating instrument for anxiety disorders. *Psychosomatics*, *12*(6), 371–9.